Iloprost reduces colitis induced oxidative stress: An experimental study in rats

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Background/aims: Reactive oxygen species have a known potent role in the pathogenesis of ulcerative colitis. Iloprost, a pharmaceutical, is a chemically stable derivative of a naturally-occurring human prostacyclin. Several studies have demonstrated protective effects of iloprost via its antioxidant and its anti-inflammatory activity. The aim of this study is to evaluate the effects of iloprost on antioxidant/antioxidant status, as well as the large bowel histopathology in experimental colitis. Materials and Method: Forty adult male Wistar-albino rats were randomly divided into four equal weight-matched groups: sham group (n=10), iloprost administered sham group (n=10), colitis group (n=10), iloprost administered colitis group (n=10). Acetic acid (1 ml of 4% solution) was used to induce colonic inflammation in the rats. Results: Colonic tissue and plasma malondialdehyde levels were significantly lower in the iloprost administered colitis group than the colitis group (p<0.01). Tissue glutathione levels of the iloprost administered colitis group were significantly higher than the colitis group (p<0.001). Conclusion: We have demonstrated in this study iloprost to be an antioxidant, as well as iloprost demonstrating protective activity against colitis induced oxidative stress.

Key words: Iloprost, prostacyclin, colitis, experimental, oxidative stress

INTRODUCTION

Reactive oxygen species (ROS) including superoxide and hydroxyl free radicals, together with hydrogen peroxide, show direct toxicity to organs. ROS can initiate free-radical-mediated chain reactions which can cause additional cellular damage (1). Malondialdehyde (MDA) is one of the end products of lipid peroxidation. Tissue concentrations of MDA are assayed as an index of membrane oxidative damage (2). Antioxidant defense mechanisms lessen the harmful effects of ROS in normal
physiological conditions. Glutathione (GSH) and superoxide dismutase (SOD) are the major known defenders of cells from oxidative damage. The oxidized form of GSH is a dimer—GSSG, and it is involved in the transport of certain amino acid. GSSH is a coenzyme for various enzymes and protects against oxygen radicals as well as other toxic compounds. SOD catalyzes the dismutation of superoxide to hydrogen peroxide. This causes the conversion of two $\text{O}_2^-$ molecules into $\text{H}_2\text{O}_2$ and $\text{O}_2$.

Increased oxidation activity is known to take place within the pathophysiology of the ulcerative colitis (UC). It is also known that ROS are involved in the development of experimental colitis (4). Local and systemic inflammation depletes the antioxidant response in UC (5).

Iloprost is a chemically stable derivative of a naturally occurring prostacyclin. Prostacyclin, which is derived from endothelium, is a known vasodilator, and it has anti thrombotic effects, inhibits activation of neutrophil leukocytes, and activates fibrinolysis (6). Several studies have demonstrated protective effects of iloprost via its antioxidant and anti-inflammatory activity (7, 8).

In literature, the data regarding iloprost against colitis is limited. The aim of this study is to evaluate the effects of iloprost on a basis of oxidant/antioxidant status, as well as the large bowel histopathology in experimental colitis.

**MATERIALS and METHODS**

This study was performed after approval from the Ethics Committee of the Animal Care Review Board of Istanbul University, Cerrahpasa Medical Faculty. Forty adult male Wistar-albino rats weighing between 200 grams and 250 grams were obtained from the Experimental Animal Production and Research Laboratory of Istanbul University Cerrahpasa Medical Faculty. These rats were randomly divided into four equal weight-matched groups: sham (S) group (n=10), after sham operation iloprost administered (IS) group (n=10), colitis (C) group (n=10), after colitis induction iloprost administered (IC) group (n=10). The rats were kept in standard colony cages (15 cm by 25 cm by 40 cm) under controlled conditions including temperature (18 °C), light (12 hours of light and then 12 hours of darkness), and controlled humidity (50-55%). The rats were fed with standard rat chow and tap water ad libitum during the experimental procedure except during the 24 - hour – period prior to the colitis induction.

**Induction of colitis**

Six centimeters of a soft six French catheter was inserted in to the anal canal of the rat, under ketamine chloride [40 mg/kg, intraperitoneal (IP)] anesthesia. Acetic acid (AA) (1 ml of 4% solution) was used for the induction of colonic inflammation. After AA injection, two ml of air was insufflated into the colon to spread the AA completely prior to the removal of the catheter. This procedure was performed for the C and the IC groups. The S and the IS groups received the same volume of isotonic saline, under the same conditions as described for the C groups.

**Iloprost administration**

The IS and the IC groups received IP 2 μg/kg/day iloprost (Ilomedin®: Bayer Schering Pharma; Bayer Turk Kimya San. Ltd. Sti, Umranie, Istanbul) immediately after colitis induction, for five consecutive days. The same volume of isotonic saline was given intraperitoneally to the S and the C groups at the same time intervals. The rats were subsequently killed by cervical dislocation five days after colitis induction, and the blood and tissue samples were then collected and processed.

**Biochemical procedures**

**Preparation of the tissue samples**

The colonic samples were diluted to 20% w/v in 20 mM ice-cold Tris–HCl, at a pH of 7.4, and homogenized with a Bosch Scintilla SA (Switzerland). The homogenate was then centrifuged at 5000 xg for 10 min, and various analytical determinations were performed in the supernatant fraction.

**Assay of malondialdehyde**

Lipid peroxidation levels in plasma and colonic tissue were measured with the thiobarbituric acid (TBA) reaction (9). This method was used to obtain a spectrophotometric measurement of the color produced during the reaction of TBA with malondialdehyde (MDA) at 535 nm. The coefficients of intra-assay and inter-assay variations for the MDA assay were 3.6% and 5.3%, respectively.

**Assay of glutathione**

The glutathione (GSH) concentrations within the colonic tissue were estimated according to the Ellman method (10). One milliliter of tissue homogenate was de-proteinized, and then subsequently centrifuged at 600 xg for 20 min. After addition of
dithiobis-nitrobenzoate and a phosphate buffer (pH 8.0) into the clear supernatants of the samples, the color this reaction produced was read at 412 nm. The GSH concentrations of the samples were calculated using 1.36 x 10^4 M cm^-1 as the molar absorption coefficient. The intra-assay and inter-assay coefficients of variation for GSH were 3.5% and 3.7%, respectively.

**Assay of Cu-Zn superoxide dismutase activity**

The plasma and the colonic tissue Cu-Zn superoxide dismutase (SOD) activity were determined utilizing the method of Sun et al. (11). The assay involves inhibition of nitroblue tetrazolium (NBT) (Sigma Chemical Co., St Louis, MO, USA) and reduction with xanthine–xanthine oxidase (Sigma Chemical Co.), which is used as a superoxide generator. One unit of SOD is defined as the amount of protein that inhibits the rate of NBT reduction by 50%. The coefficients of intra-assay and inter-assay variations were 1.8% and 3.2%, respectively.

**Histopathology evaluation**

**Macroscopic evaluation**

The large bowel of the rats were taken out in toto (from ileocecal junction to the anus), and cleaned with normal saline. Colon weight (g), colon length (cm), stool, and macroscopic inflammation were the parameters of the macroscopic evaluation. The parameters were scored from normal findings of 0 (least severe pathologic change) to 4 (the most severe pathological change) in terms of their rational change, according to the normal values. Separate index scores for these macroscopic parameters were determined for each rat.

**Microscopic evaluation**

Four randomly selected tissue sections from the distal colon were taken from the first to fourth centimeter of the anus in each rat, and were rinsed in saline, fixed in 10% neutral buffered formalin, embedded in paraffin, sectioned, and finally stained with hematoxylin and eosin (H&E). The tissues were examined and were then scored, between 0 to 3 for epithelial damage, cellular infiltration, and damage or alteration of smooth muscle architecture.

Epithelial damage was assessed in terms of blunted mucosa, destruction of crypt architecture, and/or loss of epithelium. Muscle damage was assessed in terms of sub mucosal edema, hyperplasia, and/or loss of architecture. Normal appearance was defined as 0 and severe damage (greater than 66% of tissue length) was numbered as 3. Separate index scores for microscopic parameters were determined for each rat. All of the above procedures were performed in every rat for the experimental groups. The metric scoring system for macroscopic and microscopic histopathology has been defined by Kimball et al. (12) previously.

**Statistical analysis**

All data are expressed as means, standard deviations (means±SD), and 95% confidence intervals. One-way ANOVA and post hoc Tukey’s test was used for statistical analysis, and p<0.05 was considered to be significant for all biochemical parameters.

**RESULTS**

The results of the oxidative stress parameters are summarized in Table 1. We did not observe any harmful effects of iloprost on oxidative stress parameters, macroscopic, or on microscopy of the colonic tissue. The colonic tissue (p<0.01) and plasma (p<0.001) MDA levels of the C group was determined to be significantly higher than the S group. The colonic tissue and plasma Cu-Zn SOD levels, as well as the colonic tissue GSH levels of the C group were significantly lower than the S group.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Tissue MDA</th>
<th>Tissue SOD</th>
<th>Tissue GSH</th>
<th>Plasma MDA</th>
<th>Plasma SOD</th>
</tr>
</thead>
<tbody>
<tr>
<td>S</td>
<td>1.21±0.32</td>
<td>66.89±7.96</td>
<td>5.50±1.07</td>
<td>2.73±0.78</td>
<td>22.56±2.79</td>
</tr>
<tr>
<td>IS*</td>
<td>0.96±0.26</td>
<td>68.20±6.53</td>
<td>5.34±1.10</td>
<td>2.43±0.57</td>
<td>24.70±2.67</td>
</tr>
<tr>
<td>C</td>
<td>1.85±0.52</td>
<td>53.90±9.56</td>
<td>4.02±0.54</td>
<td>4.41±0.52</td>
<td>16.90±2.38</td>
</tr>
<tr>
<td>IC</td>
<td>1.23±0.33</td>
<td>58.90±6.51</td>
<td>5.93±0.46</td>
<td>3.46±0.48</td>
<td>19.70±2.71</td>
</tr>
</tbody>
</table>

S: Sham group; IS: iloprost administered sham group; C: Colitis group; IC: iloprost administered colitis group; MDA: malondialdehyde; SOD: superoxide dismutase; GSH: glutathione.

(*) There was no difference between the biochemical parameters of the S and the IS groups. (**) Significantly different than S group (p<0.01). (†) Significantly different than S group (p<0.001). (‡) Significantly different than C group (p<0.01). ($) Significantly different than C group (p<0.001).
Iloprost administration in experimental colitis (p<0.001). The colonic tissue as well as plasma MDA levels were significantly lower in the IC group than the C group (p<0.01). Tissue GSH levels of the IC group were noted to be significantly higher than the C group (p<0.001).

Active inflammation was observed in the C group in macroscopic and microscopic analysis (Figure 1B) compared to the S group (Figure 1A).

There was no significant difference between the groups for the colon weights (p>0.05). However, the colon length of the IC group was significantly shorter than the S and IS groups (p<0.001). In the macroscopic evaluation, the stool scores of the IC group were significantly lower than the C group (p<0.01) (Table 2). In the microscopic evaluation, the muscular damage score in the IC (Figure 1C) group was significantly lower than the C group (p<0.05) (Table 3).

DISCUSSION

In this study, we have demonstrated antioxidant as well as protective activity of iloprost against oxidative stress induced by experimental colitis. Recurrent diarrhea and severe destruction of the colonic mucosa is seen almost always in the clinical presentation of UC. Unfortunately at the current time, resection of the colon and rectum is the only curative treatment option for patients. However, most cases with limited colonic extent coupled with determination of mild to moderate inflammatory activity are managed by medical treatment, generally with successful outcomes under endoscopic surveillance (13).

This reality forces physicians to find alternative medical treatment strategies for UC, and treatment which generally will continue for life (13, 14). There are various theories that attempt to

Table 2. Macroscopic scores of the experimental groups.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Colon length</th>
<th>Colon weight</th>
<th>Stools</th>
<th>Inflammation</th>
</tr>
</thead>
<tbody>
<tr>
<td>S</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>IS*</td>
<td>0.33±0.52</td>
<td>0.5±0.55</td>
<td>0.5±0.55</td>
<td>1.17±0.41</td>
</tr>
<tr>
<td>C</td>
<td>0.83±0.75</td>
<td>1±0.63</td>
<td>2.33±0.32 †</td>
<td>2.17±0.75†</td>
</tr>
<tr>
<td>IC</td>
<td>2.17±0.75†</td>
<td>1±0.63</td>
<td>1.33±0.32 §</td>
<td>1.67±0.82</td>
</tr>
</tbody>
</table>

S: Sham group; IS: iloprost administered sham group; C: Colitis group; IC: iloprost administered colitis group.

(*) There was no difference between the biochemical parameters of the S and the IS groups. (†) Significantly different than S group: p<0.001. (‡) Significantly different than C group (p<0.05). (§) Significantly different than C group (p<0.01).

Table 3. Microscopic scores of the experimental groups.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Epithelial damage</th>
<th>Cellular infiltration</th>
<th>Muscle damage</th>
</tr>
</thead>
<tbody>
<tr>
<td>S*</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>IS</td>
<td>0.33±0.52</td>
<td>0.5±0.55</td>
<td>0</td>
</tr>
<tr>
<td>C</td>
<td>3.17±0.75†</td>
<td>2.83±0.75†</td>
<td>2.33±0.52 †</td>
</tr>
<tr>
<td>IC</td>
<td>2.83±0.75</td>
<td>2.5±0.84</td>
<td>1.67±0.52 ‡</td>
</tr>
</tbody>
</table>

S: Sham group; IS: iloprost administered sham group; C: Colitis group; IC: iloprost administered colitis group.

(*) There was no difference between the biochemical parameters of the S and the IS groups. (†) Significantly different than S group: p<0.001. (‡) Significantly different than C group (p<0.05).
explain the pathophysiology of UC. One of these theories is the radical induction theory, in which uncontrolled oxidant production within the colonic epithelial cells via a pathologic cellular pathway is blamed for damaging the epithelial barrier in the theory. This initial occurrence could diminish the immune mechanisms in luminal surfaces of the large bowel, and ultimately could cause an inflammatory disease on colonic epithelium (14).

ROS have a potent role in the pathogenesis of UC. Neutrophils and macrophages produce large amounts of ROS by infiltrating the bowel wall in colitis (15). Impaired antioxidant defense in the red blood cells of UC patients have been reported (16). A tragic experience showed us the relationship between oxidants and UC in which H$_2$O$_2$ enemas had been used for the treatment of fecal impaction at the beginning of 20$^{th}$ century. With this form of enema, severe rectitis and colitis was reported following H$_2$O$_2$ enema application (13,14,16,17). H$_2$O$_2$ is also produced after aerobic and anaerobic metabolism of the cell, and its overproduction plays a key factor of many pathological pathways (3). It has been shown previously that UC causes DNA damage by increasing oxidative stress (17, 18). Colorectal cancer in UC patients occurs when there are diseased colonic epithelial cells with damaged DNA (19).

Prostacyclin regulates cAMP levels in the cell, and this controls vascular function, inhibits platelet aggregation, down-regulates neutrophil adhesion, and scavenges free radicals (20). Epithelial and stromal cells produce prostaglandins, which are important factors for cell to cell interactions in colonic tissue (21). Prostacyclin is one of the most important defenders of the gastrointestinal mucosal surfaces, as well as for the vascular structures (22). The molecular mechanism of prostacyclin activity is currently under investigation. As a stable prostacyclin analogue, the primary effect of iloprost is vasodilation. Antioxidant and anti-inflammatory properties of iloprost have been reported by presenting different molecular mechanisms on different types of tissue (6-8). The efficacy of iloprost against active colitis has not been evaluated up until now however. In our study, iloprost application augmented GSH production, which was diminished after the induction of colitis in the colonic wall. GSH is one of the body’s scavengers of H$_2$O$_2$ (22,23). Iloprost may show its antioxidant properties by initiating GSH production in colonic cells. Additionally, iloprost administration seemingly initiated a conspicuous decrease on the lipid peroxidation of the colonic tissue, as well as for the plasma. Stool formation improved in iloprost treated rats, however no significant macroscopic change were observed on the colon tissue. In the microscopic analysis, the muscular layer of the iloprost treated group appeared more natural as well as healthier than the colitis group. The beneficial effects of the same dose and application route of iloprost that we applied in the study had also been reported in previous studies investigating colon ischemia and anastomotic healing in Wistar albino rats (24,25).

While our study was being performed, Karatepe et al. (24) reported that iloprost ameliorates ischemic colitis in rats. This study presented an original idea and beneficial results were well documented in the study, however the difference of the method from the ischemia model of colon was not well defined (23,24). Ischemic colitis is difficult to evaluate experimentally, as ischemia is only one part of the complex pathogenesis of ischemic colitis (25, 26). The parameter for which they evaluated utilizing microscopy could have been changed in part to the actual ischemia, rather than as a result of colitis. This fact makes this study difficult to differentiate from a classical ischemia type presentation, versus a colitis presentation. The AA induced colitis model resembles UC in humans in terms of the histopathological appearance, due in part because of the over production of oxidant radicals (26,27). Most UC patients have rectal involvement, and UC also tends to occur on the rectal mucosa initially, and subsequently progresses retrograde to the other parts of the colon. These features of the UC are simulated with the AA induced colitis experimentally by chemical irritation of the mucosa of the large intestine from the rectum, which then extends in a retrograde fashion. Iloprost has antioxidant effects in experimental colitis and could potentially protect colon tissue against inflammation, however further studies containing inflammatory parameters are needed to better understand the effects of iloprost against colonic inflammation.
REFERENCES


